

3. M. NENNER, Dissertation, Göttingen (1970).
4. J. H. KEIJER and G. Z. WOLRING, *Biochim. biophys. Acta* **185**, 465 (1969).
5. A. J. J. OOMS and C. VAN DIJK, *Biochem. Pharmac.* **15**, 1361 (1966).
6. M. L. BENDER, G. R. SCHONBAUM and B. ZERNER, *J. Am. Chem. Soc.* **84**, 2540 (1962).
7. K. SCHOENE, to be published.

Biochemical Pharmacology, Vol. 20, pp. 2529-2531. Pergamon Press, 1971. Printed in Great Britain

Metabolism of diazepam and its metabolites by guinea pig liver microsomes

(Received 15 February 1971; accepted 25 March 1971)

LIVER microsomal preparations of various animal species metabolize diazepam added *in vitro*. This drug is predominantly hydroxylated in C₃ position and only slightly *N*-demethylated by rat liver¹ while it is more *N*-demethylated than hydroxylated in mice.¹ Previous studies *in vivo* indicated that guinea pigs are also able to metabolize diazepam.² This note summarizes the findings obtained by adding diazepam and some of its known metabolites (see Fig. 1) to guinea pig liver microsomes.

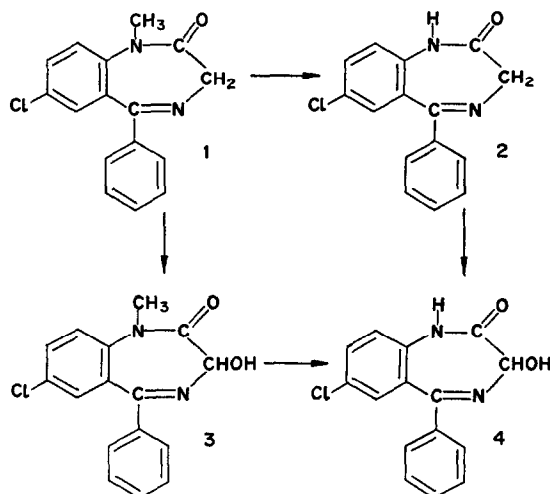


FIG. 1. (1) Diazepam. (2) *N*-demethyldiazepam. (3) *N*-methyloxazepam. (4) Oxazepam.

Materials and Methods

(1) *Animals*. Male Albino guinea pigs (body weight 300–350 g) fed with Alcom-Alal pellets, were used in all experiments.

(2) *Microsome preparation*. Animals were killed and the livers were immediately removed and homogenized in ice-cold 1.15% KCl solution (1:4 w/v) with a teflon glass homogenizer. The homogenate was centrifuged at 9000 *g* for 20 min and then the supernatant fraction was again centrifuged at 105,000 *g* for 1 hr (rotor 40'—Beckman Model L ultracentrifuge). The liver microsomes were suspended in 1.15% KCl solution.

(3) *Incubation in vitro*. The incubation mixture contained substrate, enzyme source, pyridine nucleotides and an NADPH-generating system according to that described by Kato *et al.*³ The incubation volume of 5 ml consisted of 2.5 ml of microsomal suspension equivalent to 1 g of liver,

TABLE 1. *In vitro* METABOLISM OF DIAZEPAM AND DIAZEPAM METABOLITES BY GUINEA PIG LIVER MICROSOMES

Drug added (nmoles)	Drugs after incubation (nmoles \pm S.E.)				
	Diazepam	N-Methyl- oxazepam	N-Demethyl- diazepam	Oxazepam	Recovery*
Diazepam 35	0.016	†	32 \pm 0.45		91.4
70	0.016		65 \pm 0.29		92.8
105	37 \pm 0.1		69 \pm 0.1		100.9
140	62 \pm 0.7		80 \pm 0.7		101.4
210	76 \pm 0.7		130 \pm 3.6		98.0
350	134 \pm 0.3		187 \pm 4.4		91.7
525	243 \pm 0.7		254 \pm 1		94.6
700	395 \pm 0.3		284 \pm 3.6		97.0
1050	754 \pm 0.7		309 \pm 3.6		101.2
1700	1471 \pm 4.2		295 \pm 4.4		100.9
N-methyl- oxazepam 33				7 \pm 0.3	21.2
66				16 \pm 1.0	24.2
99		1.6		20 \pm 0.6	20.2
199		126 \pm 3.9		35 \pm 3.8	80.9
332		274 \pm 4.9		24 \pm 1.7	89.7
664		623 \pm 10.2		17 \pm 1.0	96.3
1661		1641 \pm 16.6		10 \pm 0.6	99.3
N-demethyl- diazepam 36			32 \pm 1.1		88.8
73			73 \pm 3.6		100.0
110			102 \pm 2.9		92.7
220			215 \pm 3.6		97.7
367			361 \pm 4.0		98.3
735			735 \pm 4.7		100.0
1830			1823 \pm 5.5		99.6
Oxazepam 34				31 \pm 0.3	91.0
69				65 \pm 1.2	94.2
104				102 \pm 1.8	98.0
209				207 \pm 2.1	99.0
349				346 \pm 3.6	99.1
699				601 \pm 3.6	85.9
1748				1730 \pm 4.2	98.9

* These figures represent the sum of diazepam and its metabolites found after the incubation. The values are expressed as per cent of the added diazepam or diazepam metabolite.

† When there is a blank space the compounds were not present. (The limit of sensitivity of the method was 0.05 μ g.)

NADP (1.5 μ moles), glucose-6-phosphate (50 μ moles), glucose-6-phosphate dehydrogenase (0.5 units), magnesium chloride (25 μ moles), nicotinamide (50 μ moles), 1.4 ml of 0.2 M phosphate buffer pH 7.4, the substrate (diazepam; N-demethyldiazepam; N-methyloxazepam or oxazepam) dissolved in ethanol and added in amounts ranging from 10 to 500 μ g and 0.45 ml of 1.15% KCl.

The incubation was carried out at 37° for 10 min under air in a Dubnoff metabolic shaker.

(4) *Chemical determinations.* After the incubation the entire content of each flask was extracted twice with 10 ml of fresh peroxide-free ethylether. The combined extracts were evaporated to dryness, redissolved in a suitable amount of acetonitrile and then gas-chromatographed according to the method previously reported.^{4,5}

Results and discussion

Table 1 summarizes the findings obtained by incubating *in vitro* guinea pig liver microsomes with diazepam and three among its known metabolites. It appears that only N-demethylated products

accumulate in these experimental conditions. In fact *N*-demethyldiazepam and oxazepam are obtained after incubating diazepam and *N*-methyloxazepam respectively, while *N*-demethyldiazepam and oxazepam are not furtherly metabolized and they are almost completely recovered as such. The rate of *N*-demethylation of diazepam at saturating concentrations is calculated as 1854 nmoles/g of tissue/hr. In the case of low concentrations of *N*-methyloxazepam the recovered amount of the unmetabolized product added to the respective formed metabolite does not account for the added substrate. Since unpublished studies showed a recovery of *N*-methyloxazepam added to guinea pig liver microsomes of about 90 per cent (the presented data are corrected for the respective recovery factor), the noticed loss of this substrate may suggest a possible alternate metabolic pathway which leads to the formation of unknown compounds.

The findings obtained for diazepam and *N*-demethyldiazepam are consistent with the *in vivo* studies showing that no C₃-hydroxylated metabolites accumulate in blood brain and adipose tissue.² It is interesting to notice that diazepam is metabolized by guinea pigs both *in vitro* and *in vivo*² in a different manner than by rats or mice. As far as the *in vitro* metabolism is concerned, *N*-demethyldiazepam is slightly hydroxylated by rats and mice but not by guinea pigs while *N*-methyloxazepam is demethylated by mice and guinea pigs but not by rats. Oxazepam is not further metabolized *in vitro* by the liver microsomes of the three animal species here considered.¹

Acknowledgement—This work was supported by NIH Contract No. DHEW/PHS.NIH/PH 43-67-83. The technical assistance of Mr. Claudio Reschiotto is gratefully acknowledged.

Istituto di Ricerche Farmacologiche, Mario 'Negri',
Via Eritrea 62,
20157 Milan,
Italy

E. MUSSINI
F. MARCUCCI
R. FANELLI
S. GARATTINI

REFERENCES

1. F. MARCUCCI, R. FANELLI, E. MUSSINI and S. GARATTINI, *Europ. J. Pharmac.* **7**, 307 (1969).
2. F. MARCUCCI, A. GUAITANI, R. FANELLI, E. MUSSINI and S. GARATTINI, to be published in *Europ. J. Pharmac.* (1970).
3. R. KATO and M. TAKAYANAGHI, *Jap. J. Pharmac.* **16**, 380 (1966).
4. S. GARATTINI, F. MARCUCCI and E. MUSSINI, in *Gas Chromatography in Biology and Medicine, a Ciba Foundation Symposium* (Ed. R. PORTER), p. 161. Churchill, London (1969).
5. F. MARCUCCI, R. FANELLI and E. MUSSINI, *J. Chromat.* **37**, 318 (1968).

Biochemical Pharmacology, Vol. 20, pp. 2531-2534. Pergamon Press, 1971. Printed in Great Britain

Nachweis der 16-Hydroxylierung bei Pregnanen in Ratten nach Barbiturat-Gabe durch Radiospirometrie

(Received 3 December 1970; accepted 25 March 1971)

DAS PRINZIP der Stoffwechsel-labilen Tritium-Markierung verwendeten wir zur *in-vivo* Messung der Hydroxylierung bereits Hydroxylgruppen tragender Pregnanderivate bei Ratten. Untersucht wurden die folgenden Steroide,* die alle in Position 16 stabil mit Tritium markiert waren¹

3 α , 17 α , 20 β -Trihydroxy-5 β -pregnan (Trihydroxypregnan)
3 α , 17 α -Dihydroxy-20-oxo-5 β -pregnan (Dihydroxypregnan)
3 α -Hydroxy-20-oxo-5 β -pregnan (Pregnanolon)

* Für die Überlassung der Steroide haben wir Herrn Dr. J. Livet, Paris, zu danken (vergl. 1).